

Nitrate and nitrite in biology, nutrition and therapeutics

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Inorganic nitrate and nitrite from endogenous or dietary sources are metabolized *in vivo* to nitric oxide (NO) and other bioactive nitrogen oxides. The nitrate-nitrite-NO pathway is emerging as an important mediator of blood flow regulation, cell signaling, energetics and tissue responses to hypoxia. The latest advances in our understanding of the biochemistry, physiology and therapeutics of nitrate, nitrite and NO were discussed during a recent 2-day meeting at the Nobel Forum, Karolinska Institutet in Stockholm.

Until recently the inorganic anions nitrate (NO_3^-) and nitrite (NO_2^-) were considered inert end products of nitric oxide (NO) metabolism and unfavorable dietary constituents. However, a new view is evolving with the accumulating evidence that nitrate and nitrite metabolism occurs in blood and tissues to form NO and other bioactive nitrogen oxides, representing an alternative to 'classical' L-arginine-NO synthase-NO signaling¹⁻⁵. In situations of hypoxia, when the oxygen-dependent NO synthases may become dysfunctional, nitrite reduction is instead greatly enhanced. Nitrate and nitrite can thus be viewed as storage pools supporting NO signaling during metabolic stress, with their bioactivation involving both enzymatic and nonenzymatic reactions in blood and tissues. The bioactivation of the more stable nitrate anion involves initial reduction to nitrite by symbiotic bacteria in the oral cavity¹. A role for nitrite in regulation of blood flow and tissue responses to hypoxia has been reported. In addition, multiple studies now support a therapeutic role for nitrate and nitrite, most notably in the treatment and prevention of cardiovascular diseases including ischemia-reperfusion (IR) injury and hypertension¹. The nutritional aspects of these cardioprotective effects are particularly intriguing since nitrate and nitrite are abundant in our everyday diet. On June 17th

and 18th, 2009, a 2-day symposium was held at the Nobel Forum, Karolinska Institutet in Stockholm, Sweden to discuss recent advances in understanding the role of nitrate and nitrite in biology, nutrition and therapeutics.

Nitric oxide, nitrite and mitochondria

Recent data suggest that many of the biological effects of nitrite^{6,7} involve interaction with mitochondria. In the last 15 years it has been established that the mitochondrion is a physiological target for NO (ref. 8). In the opening

keynote lecture, Salvador Moncada (University College London) gave an overview on how NO regulates mitochondrial function in health and disease. NO binds to cytochrome *c* oxidase, the terminal electron acceptor in the mitochondrial electron transport chain, in competition with oxygen. Moncada reviewed evidence showing that binding of NO to cytochrome *c* oxidase elicits intracellular signaling events, including the diversion of oxygen to non-respiratory substrates and the generation of reactive oxygen species (ROS) with potentially



The Nobel Forum at Karolinska Institutet, where the Third International Meeting on the Role of Nitrate and Nitrite in Physiology, Pathophysiology and Therapeutics was held.

See end for affiliations.

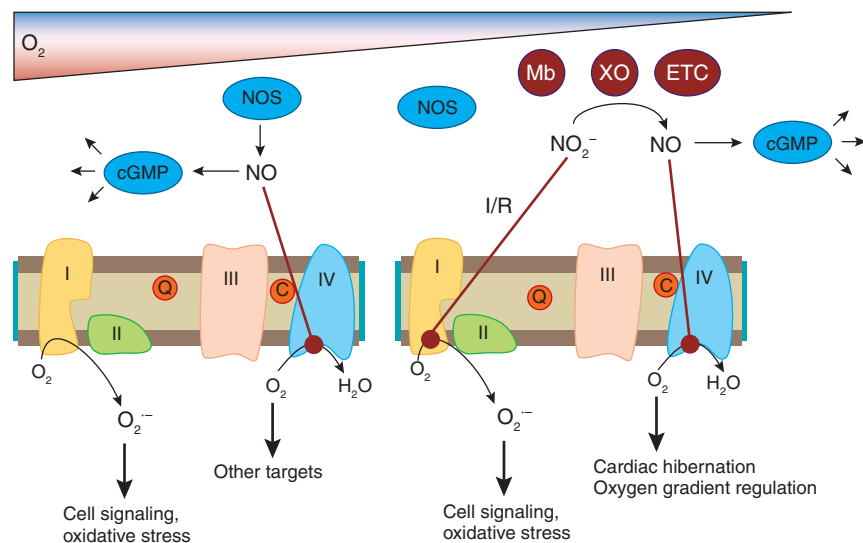


Figure 1 Nitric oxide and nitrite interactions with mitochondria. In normoxic conditions NOS generates NO, which activates cGMP signaling pathways (for example, vasodilation) and directly inhibits complex IV. This allows oxygen to be diverted away from the mitochondria to other targets (such as HIF1 α). This inhibition also increases ROS generation from the mitochondria, generating increased ROS to potentially mediate cell signaling and oxidative stress. In hypoxia/ischemia, NOS activity is inhibited by the lack of oxygen. Nitrite is reduced to NO by deoxygenated myoglobin (Mb), xanthine oxidoreductase (XO), enzymes of the electron transport chain (ETC) or other enzymes, and the resulting NO can inhibit respiration at complex IV to regulate oxygen gradients or mediate cardiac hibernation. During ischemia/reperfusion, nitrite inhibits complex I via S-nitrosation, which decreases ROS generation at reperfusion, leading to cytoprotection.

damaging effects. He also discussed findings indicating that these NO-elicited events act as triggers by which mitochondria modulate signal transduction cascades involved in the induction of cellular defense mechanisms and adaptive responses. Furthermore, he presented evidence indicating that lack of NO metabolism by cytochrome *c* oxidase at low oxygen concentrations might explain the phenomenon of hypoxic vasodilatation.

Sruti Shiva (University of Pittsburgh) reviewed a number of studies demonstrating that components of the respiratory chain can reduce nitrite to NO (refs. 4,9), as well as data suggesting that nitrite reduction by myoglobin and xanthine oxidoreductase (XOR) can generate NO during hypoxia that then regulates mitochondrial respiration (Fig. 1). Although the physiological relevance of this is still uncertain, recent data suggest that nitrite-dependent modulation of mitochondrial function may be important in regulating oxygen gradients during hypoxia and mediating cytoprotection after IR injury⁶ (see below).

The biochemistry and bioactivation of nitrite

Bacteria commonly utilize nitrate and nitrite as terminal electron acceptors for respiration in the absence of oxygen, or for incorporation in biomass. However, it was only recently described

that mammalian tissues can also reduce nitrate¹⁰ and nitrite¹¹ to bioactive nitrogen oxides including NO (Fig. 2). It is now clear that the one-electron reduction of nitrite to NO can be catalyzed by numerous enzymatic as well as nonenzymatic pathways¹⁻⁴. The relative contribution from these pathways to overall NO formation varies between tissues and is dependent on a number of factors including local pH, oxygen tension and redox status.

The ability of deoxygenated hemoglobin and myoglobin to react with and reduce nitrite to NO is currently a matter of intense research, which was reflected by the many presentations on this topic. Tienush Rassaf (University of Dusseldorf) explained that myoglobin-derived NO inhibits cellular respiration and limits ROS formation and mitochondrial enzyme dysfunction after IR injury^{6,12}. Nitrite protection against myocardial infarction is lacking in myoglobin knockout mice, which supports the hypothesis that myoglobin serves a critical function as an intrinsic nitrite reductase that regulates cellular responses to hypoxia and reoxygenation¹³. The family of nitrite-reducing globins has been expanded to include neuroglobin—a monomeric heme protein specifically expressed in the brain and the most ancestral among vertebrate globins^{14,15}. Mark Gladwin (University of Pittsburgh) explained that neuroglobin, unlike hemoglobin and

myoglobin, has a six-coordinate heme geometry. Such heme geometry greatly limits oxygen binding, raising questions about the physiological functions of these six-coordinate globins. Nevertheless, the expression of neuroglobin is associated with cytoprotective effects in various animal models of brain ischemia. Gladwin showed that if its five-coordinate state is made stable, neuroglobin functions as a very efficient nitrite reductase. He speculated that six-coordinate heme globins in general, including neuroglobin, may function as post-translationally (via a six- to five-coordinate equilibrium) regulated nitrite reductases and primordial NO synthases whose function antedates the evolution of oxygen carriers.

Although many of the biological effects of nitrite may proceed via direct reduction to NO, other bioactive species can also be formed. In addition, it has been suggested that nitrite can signal directly, possibly via nitrosation of thiols and nitrosylation of hemes, without intermediate NO formation¹⁶. Jack Lancaster (University of Alabama at Birmingham) explained an interesting potential pathway for the generation of nitrosothiols involving NO, nitrate and nitrite reactions with nonheme iron to form dinitrosyl iron complexes¹⁷. The nitrite anion is also involved in nitration reactions that are commonly implicated in pathological processes such as inflammation and atherosclerosis. Bruce Freeman (University of Pittsburgh) reviewed data on the endogenous formation of nitrated unsaturated fatty acids (NO₂-FA) that in turn mediate adaptive and anti-inflammatory signaling reactions. Fatty acid nitration is induced by NO and nitrite-derived species reacting via multiple mechanisms, ultimately involving homolytic addition of nitrogen dioxide (NO₂) to the double bond. NO₂-FAs display cGMP-independent signaling activity as a consequence of robust electrophilic reactivity, facilitating reversible adduction to nucleophilic targets such as cysteine residues¹⁸. Overall, these presentations highlighted the function of nitrite as a relatively stable reservoir of NO bioactivity, which can also undergo secondary reactions with biomolecules to form redox-derived signaling mediators.

Nitrite reduction in the vasculature

NO-mediated control of blood flow involves direct diffusion of NO from the primary site of generation in the endothelium. In addition, the red blood cell (RBC) contributes to vasoregulation by exporting NO-like bioactivity as discussed by Jonathan Stamler (Duke University), who originally proposed this mechanism¹⁹. Different mechanisms for the generation and export of NO bioactivity from the RBC have been proposed^{14,19,20}. At this meeting the role

of nitrite as a source of NO in the circulation was highlighted. Nitrite administration causes vasodilatation at low concentrations¹⁴, and this is ultimately mediated by activation of cGMP in the vascular smooth muscle. The sites of production and the mechanism for nitrite-dependent NO generation in the vasculature were vigorously discussed, and it was clear that there is still controversy about the significance of endogenous nitrite in physiological regulation of blood vessel tone. Deoxygenated hemoglobin has been implicated in controlling nitrite-dependent NO signaling in the vasculature^{14,20}. Rakesh Patel (University of Alabama at Birmingham) discussed the potential mechanisms that may regulate nitrite-RBC interactions, including oxygen fractional saturation-dependent control over nitrite transport into the RBC and heme-based nitrite reduction²¹. Nitrite reduction to NO by deoxygenated hemoglobin may govern NO signaling in the vasculature as exemplified by experiments showing that nitrite therapy can prevent hypertension induced by cell-free hemoglobin-based oxygen carriers *in vivo*²². A remaining controversial issue is how any NO formed in the red cell is exported to elicit vasodilatation. Given the extremely fast rate that NO reacts with hemoglobin, export of free NO from the red blood cell is unlikely. Daniel Kim-Shapiro (Wake Forest University) explained recent data describing formation of the nitrosating agent N_2O_3 from the concerted nitrite reductase/anhydrase activity of hemoglobin²³. Because N_2O_3 is less reactive with hemoglobin (Hb) or HbO_2 than is NO, this could facilitate export of NO bioactivity from the RBC, either directly as N_2O_3 or via formation of a secondary S-nitrosothiol. Joel Friedman (Albert Einstein College of Medicine) explained how he has been using sol-gel and glassy matrices to dissect the nitrite-hemoglobin interactions on a molecular level²⁴. Both Friedman and Pedro Cabrales (University of California) showed data supporting a nitrite reductase and anhydrase activity of hemoglobin as well as hemoglobin-based oxygen carriers. George Richter-Addo (University of Oklahoma) demonstrated, through crystallography, that nitrite binds to the ferric iron center of the protein in an unusual O-binding mode²⁵. Interestingly, conformational differences exist in the O-binding of nitrite to the α and β subunits of the protein.

Amrita Ahluwalia (Barts and The London Medical School) presented data demonstrating expression of eNOS and XOR on the membranes of human RBCs and blood vessels²⁶. By using inhibitors of XOR and NO synthase, Ahluwalia suggested that the nitrite reductase activity within the circulation (under hypoxic conditions and physiological pH) is mediated in part by eNOS, while a substantial role for XOR

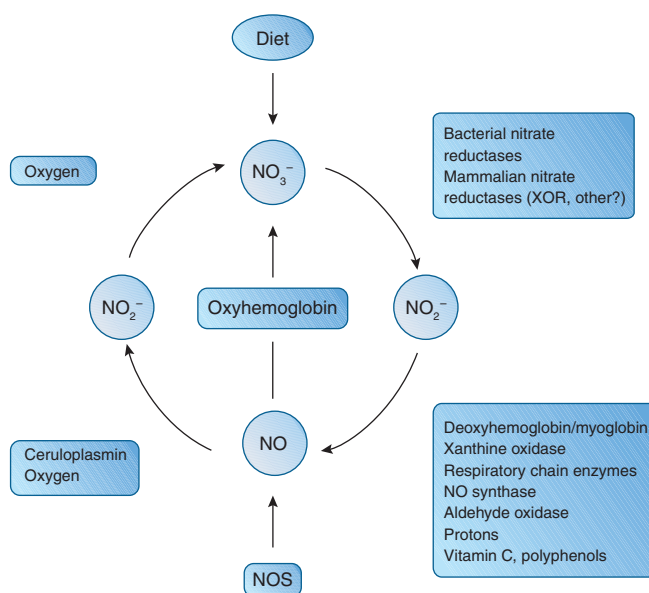


Figure 2 A mammalian nitrogen oxide cycle. Nitric oxide synthase (NOS) generates NO in cells to regulate a vast variety of physiological functions. The bioactivity of NO is acutely terminated by its rapid oxidation to nitrite and nitrate. In our bodies nitrate can undergo reduction to nitrite (NO_2^-) by bacteria in the oral cavity and by xanthine oxidase and possibly other enzymes in tissues. In blood and tissues nitrite can be further metabolized to nitric oxide (NO) and other biologically active nitrogen oxides (not shown in figure). This reduction is catalyzed by various enzymatic and nonenzymatic pathways, most of which are greatly enhanced under hypoxic conditions. The reduction of nitrate and nitrite to NO completes a mammalian nitrogen oxide cycle.

becomes evident as acidosis develops. Further studies are required to determine exactly when and where XOR is recruited to act as a nitrite reductase *in vivo*. Jay Zweier (Ohio State University) discussed the mechanisms for nitrite reduction in the vessel wall. NO formation was detected by electrochemical and electron paramagnetic resonance techniques in isolated vessels that dilated following nitrite treatment. These events could be blocked by adding either hemoglobin or an inhibitor of soluble guanylyl cyclase (sGC)²⁷. Zweier suggested that sGC or a resident heme protein might catalyze nitrite reduction to NO.

Dietary nitrate in health and disease

Our everyday diet is a major source of nitrate, and the levels are particularly high in certain vegetables. As an example, one serving of spinach, lettuce or beetroot contains more nitrate than that generated endogenously over a day by all three NOSs combined¹. A substantial amount of ingested nitrate is actively taken up from the circulation by the salivary glands and concentrated in saliva. Salivary nitrate is then efficiently reduced to nitrite by bacteria in the oral cavity, leading to 1,000-fold higher concentrations of nitrite in saliva compared to plasma²⁸. The continuous swallowing of nitrite-containing saliva (~1.5 liters per day) generates a variety of nitrogen oxides, including NO, in the acidic milieu of the gastric lumen. Actually,

the initial discoveries that nitrate and nitrite can be converted to NO in mammals came from observations that the gastric lumen contains high levels of NO (refs. 29,30). The enterosalivary circulation of nitrate also leads to a sustained increase in plasma levels of nitrite³¹, and nitrate may therefore be considered as a 'prod-rug' of nitrite with a slow-release profile.

Over the past half century, dietary nitrate and nitrite have become notorious because of their supposed association with the development of disease, most notably gastric cancer³². This has led to strict regulations of nitrate and nitrite levels in food and drinking water. Kenneth McColl (Glasgow University) described the potential harmful effects of nitrate-derived gastric NO. The generation is maximal at the gastroesophageal junction where salivary nitrite first meets gastric acid³³. The NO formed diffuses into the tissue where it forms nitrosating species, which in turn may cause DNA damage and promote carcinogenesis. According to McColl ascorbic acid would paradoxically further aggravate the carcinogenic effects since this vitamin greatly enhances NO formation from acidified nitrite. In the same session Nigel Benjamin (University of Exeter and Plymouth) surveyed the underlying epidemiological evidence for the recommended limitations of nitrate and nitrite in food and drinking water. He concluded that there is relatively weak evidence to support the present limitations³⁴, especially in light of the

beneficial effects of nitrate and nitrite that have emerged during recent years. Interestingly, a diet rich in vegetables, such as the Mediterranean and the traditional Japanese diets, contains more nitrate than that recommended as the acceptable daily intake (ADI) by the World Health Organization³⁴. Joel Petersson (Uppsala University) discussed physiological and therapeutic NO-dependent effects of dietary nitrate in the gastrointestinal tract. Salivary nitrite increases gastric mucosal blood flow and mucus production, thereby supporting gastroprotective mechanisms³⁵.

A diet rich in fruit and vegetables reduces blood pressure and protects against cardiovascular disease, but the active ingredients responsible for these effects have not been pinpointed. Recently it was shown that modest dietary supplementation with sodium nitrate reduces blood pressure in healthy volunteers³⁶. Andrew Webb (Barts and The London Medical School) and colleagues used beetroot juice as a source of nitrate in healthy volunteers and found a reduction in blood pressure of ~10 mm Hg that coincided with peak plasma levels of nitrite³⁷. In addition, inhibitory effects on *ex vivo* platelet aggregation and protection against IR-induced endothelial dysfunction were observed. Similar data were presented by Mattias Carlström (Uppsala University), who observed a sustained effect on blood pressure by chronic dietary nitrate supplementation in rats³⁸. Both groups could prevent the effects on blood pressure either by letting the human volunteers spit for 3 h or by application of an antibacterial mouthwash to the rats. This shows the importance of the enterosalivary circulation of nitrate and the critical role of oral nitrate-reducing bacteria in its bioactivation. Nathan Bryan (University of Texas) argued that nitrate and nitrite should be viewed as bioactive food components and indispensable nutrients that may serve to uphold NO/nitroso homeostasis, especially in conditions where the L-arginine-NOS pathway is dysfunctional. Bryan explained that dietary nitrate and nitrite protect against IR injury in experimental myocardial infarction³⁹; and in mice fed a cholesterol-enriched diet, the addition of nitrite to the drinking water prevented the signs of vascular pathology⁴⁰.

Nitrate, nitrite and energetics

The role of NO in regulation of mitochondrial function and energy expenditure has attracted much attention. Recently it was demonstrated that dietary nitrate supplementation reduces oxygen cost and improves muscular efficiency at various levels of submaximal exercise in healthy young volunteers without increasing anaerobic metabolism⁷. Andy Jones (University of Exeter) presented data demonstrating that

in healthy volunteers given a dietary nitrate load (in the form of beetroot juice), oxygen consumption was reduced, resulting in a concomitant improvement of performance⁴¹. The mechanisms underlying these remarkable effects of dietary nitrate need further investigation, and the discussion at the meeting focused on effects at the mitochondrial level. Similar mechanisms may be at work when nitrite and nitrate are produced from endogenous NO. To this end, Martin Feelisch (University of Warwick) presented preliminary results from an ongoing study in mountaineers, who experience an integrated whole-body response to exercise and hypoxia. While ascent to altitude was associated with enhanced oxidative stress and discrete increases in circulating nitrite and nitrate, no simple relationship of these “NO markers” to the prevailing level of hypoxia was apparent, which suggests that the *in vivo* situation is complex. It is established that exercise provides protection against cardiovascular disease, and David Lefer (Emory University School of Medicine) showed data on exercise-induced increases in circulating and myocardial nitrite and S-nitrosothiol levels in mice. The increase in myocardial storage of nitrite after exercise was associated with protection during acute myocardial infarction.

Therapeutic opportunities

In recent years data have accumulated showing that nitrite protects against IR injury in the heart, brain, kidney and liver, as well as in chronic limb ischemia (for review see refs. 1,2). At the meeting a number of new therapeutic indications were discussed. In their presentations Brian Zuckerbraun (University of Pittsburgh) and Erik Swenson (University of Washington) focused on the pulmonary effects of nitrite where both inhalation therapy and intravenous administration were shown to be protective in models of pulmonary hypertension, ventilator-induced lung injury and lung IR injury.

Renal injury is a common complication in people with hypertension. Koichiro Tsuchiya (University of Tokushima) presented data showing that very low doses of dietary nitrite attenuated proteinuria and prevented the renal histological damage induced by chronic administration of an NOS inhibitor⁴². These data indicate that the nitrate-nitrite-NO pathway can compensate for disturbances in endogenous NO generation from NOSs.

John Lang (University of Washington) and Kenneth McCurry (Cleveland Clinic) discussed ongoing studies on nitrite therapy during solid organ transplantation. They demonstrated beneficial effects of nitrite in animal models of liver and heart transplantation, respectively. Administration of nitrite to

the organ preservation fluid and to the organ recipient improved parameters of organ survival and function⁴³.

In aggregate, promising data are emerging in a wide variety of conditions predominantly involving tissue ischemia. This insight is catalyzing the evaluation of potential therapeutic benefits of nitrate and nitrite administration, in the context of L-arginine-NO signaling. A motivation for this is the limited success realized when therapeutic trials have been pursued for clinical indications using clinically prevalent NO donors and inhibitors of NO synthases. This could be due to the ubiquitous presence of different NOS isoforms, the pluri-potency of NO and the rather nonselective release of NO from most donors. In contrast, nitrite bioactivation is promoted in anatomic regions undergoing ischemia/hypoxia, leading to potentially more localized therapeutic effects. Another possible advantage is the apparent absence of tolerance connected to relatively long-term nitrite⁴⁴ or nitrate³⁸ administration in contrast to treatment with organic nitrates such as nitroglycerin.

Conclusions and future perspectives

During a rather short time span our insight into the nitrate-nitrite-NO pathway has improved dramatically. Now, future research challenges include addressing a range of critical issues. Perhaps the most important is determining the physiological roles of endogenous nitrate and nitrite levels. This is not trivial, however, since in contrast to NOS-dependent physiology, which has been largely explored by the use of NOS inhibitors, there is no specific nitrite reductase inhibitor available. Moreover, the dual origin of nitrate and nitrite (from our diet and from endogenous production) presents a vexing obstacle in experimental design.

The mechanisms accounting for nitrite bioactivation clearly will include new candidate enzymes that may be specific for different tissues and/or at different pH or oxygen gradients. Moreover, it is important to understand the mechanisms and compartments within the vasculature where nitrite reduction occurs and the roles of various physiological pH and oxygen tension extremes in the generation of various metabolites. Also, a greater appreciation is needed of how endogenous levels of nitrate and nitrite are regulated and transported and how they interface with the NOS pathway.

This report reveals new perspectives on the therapeutic use of nitrate and nitrite that are actively emerging and consequently setting the stage for future clinical trials related to conditions involving ischemia and reperfusion. The nutritional aspects of the nitrate-nitrite-NO pathway are also particularly intriguing

and will motivate a vibrant discussion challenging the dogma that dietary nitrate and nitrite are harmful. Of crucial relevance, the salutary effects of nitrate and nitrite that are emerging may also explain some of the beneficial health effects of a diet rich in vegetables, such as the Mediterranean diet, thereby instigating a discussion of the revision of acceptable levels of intake.

In summary, future laboratory, clinical and epidemiologic investigations of the role of nitrate and nitrite in physiology, pathophysiology, nutrition and therapeutics promise to bear fruit regarding the development of new therapeutic avenues for important clinical problems and can shift current views on our dietary constituents in the context of health and disease.

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COMPETING INTERESTS STATEMENT

The authors declare competing financial interests: details accompany the full-text HTML version of the paper at <http://www.nature.com/naturechemicalbiology/>.

- Lundberg, J.O., Weitzberg, E. & Gladwin, M.T. *Nat. Rev. Drug Discov.* **7**, 156–167 (2008).
- Gladwin, M.T. *et al. Nat. Chem. Biol.* **1**, 308–314 (2005).
- Reutov, V.P. & Sorokina, E.G. *Biochemistry (Mosc.)* **63**, 874–884 (1998).
- van Faassen, E.E. *et al. Med. Res. Rev.* **29**, 683–741 (2009).
- Weitzberg, E. & Lundberg, J.O. *Nitric Oxide* **2**, 1–7 (1998).
- Shiva, S. *et al. J. Exp. Med.* **204**, 2089–2102 (2007).
- Larsen, F.J., Weitzberg, E., Lundberg, J.O. & Eklom, B. *Acta Physiol. (Oxf.)* **191**, 59–66 (2007).
- Erusalimsky, J.D. & Moncada, S. *Arterioscler. Thromb. Vasc. Biol.* **27**, 2524–2531 (2007).
- Kozlov, A.V., Staniek, K. & Nohl, H. *FEBS Lett.* **454**, 127–130 (1999).
- Jansson, E.A. *et al. Nat. Chem. Biol.* **4**, 411–417 (2008).
- Zweier, J.L., Wang, P., Samouilov, A. & Kuppusamy, P. *Nat. Med.* **1**, 804–809 (1995).
- Rassaf, T. *et al. Circ. Res.* **100**, 1749–1754 (2007).
- Hendgen-Cotta, U.B. *et al. Proc. Natl. Acad. Sci. USA* **105**, 10256–10261 (2008).
- Cosby, K. *et al. Nat. Med.* **9**, 1498–1505 (2003).
- Petersen, M.G., Dewilde, S. & Fago, A. *J. Inorg. Biochem.* **102**, 1777–1782 (2008).
- Bryan, N.S. *et al. Nat. Chem. Biol.* **1**, 290–297 (2005).
- Bosworth, C.A., Toledo, J.C. Jr., Zmijewski, J.W., Li, Q. & Lancaster, J.R. Jr. *Proc. Natl. Acad. Sci. USA* **106**, 4671–4676 (2009).
- Freeman, B.A. *et al. J. Biol. Chem.* **283**, 15515–15519 (2008).
- Jia, L., Bonaventura, C., Bonaventura, J. & Stamler, J.S. *Nature* **380**, 221–226 (1996).
- Nagababu, E., Ramasamy, S., Abernethy, D.R. & Rifkind, J.M. *J. Biol. Chem.* **278**, 46349–46356 (2003).
- Vitturi, D.A. *et al. Am. J. Physiol. Heart Circ. Physiol.* **296**, H1398–H1407 (2009).
- Rodriguez, C. *et al. Biochem. J.* **422**, 423–432 (2009).
- Basu, S. *et al. Nat. Chem. Biol.* **3**, 785–794 (2007).
- Roche, C.J., Dantsker, D., Samuni, U. & Friedman, J.M. *J. Biol. Chem.* **281**, 36874–36882 (2006).
- Yi, J., Safo, M.K. & Richter-Addo, G.B. *Biochemistry* **47**, 8247–8249 (2008).
- Webb, A.J. *et al. Circ. Res.* **103**, 957–964 (2008).
- Alzawhra, W.F., Talukder, M.A., Liu, X., Samouilov, A. & Zweier, J.L. *Am. J. Physiol. Heart Circ. Physiol.* **295**, H499–H508 (2008).
- Lundberg, J.O., Weitzberg, E., Cole, J.A. & Benjamin, N. *Nat. Rev. Microbiol.* **2**, 593–602 (2004).
- Lundberg, J.O., Weitzberg, E., Lundberg, J.M. & Alving, K. *Gut* **35**, 1543–1546 (1994).
- Benjamin, N. *et al. Nature* **368**, 502 (1994).
- Lundberg, J.O. & Govoni, M. *Free Radic. Biol. Med.* **37**, 395–400 (2004).
- Tannenbaum, S.R. & Correa, P. *Nature* **317**, 675–676 (1985).
- Iijima, K. *et al. Gastroenterology* **122**, 1248–1257 (2002).
- Alexander, J. *et al. EFSA J.* **689**, 1–79 (2008).
- Bjorne, H.H. *et al. J. Clin. Invest.* **113**, 106–114 (2004).
- Larsen, F.J., Eklom, B., Sahlin, K., Lundberg, J.O. & Weitzberg, E. *N. Engl. J. Med.* **355**, 2792–2793 (2006).
- Webb, A.J. *et al. Hypertension* **51**, 784–790 (2008).
- Petersson, J. *et al. Free Radic. Biol. Med.* **46**, 1068–1075 (2009).
- Bryan, N.S. *et al. Proc. Natl. Acad. Sci. USA* **104**, 19144–19149 (2007).
- Stokes, K.Y. *et al. Am. J. Physiol. Heart Circ. Physiol.* **296**, H1281–H1288 (2009).
- Bailey, S.J. *et al. J. Appl. Physiol.* **107**, 1144–1155 (2009).
- Kanematsu, Y. *et al. Am. J. Physiol. Renal Physiol.* **295**, F1457–F1462 (2008).
- Zhan, J. *et al. Surgery* **146**, 155–165 (2009).
- Dejam, A. *et al. Circulation* **116**, 1821–1831 (2007).

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